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Intraoral digital scans-Part 1: Influence of ambient scanning light conditions on the accuracy (trueness and precision) of different intraoral scanners

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Abstract: STATEMENT OF PROBLEM Digital scans have increasingly become an alternative to conventional impressions. Although previous studies have analyzed the accuracy of the available intraoral scanners (IOSs), the effect of the light scanning conditions on the accuracy of those IOS systems remains unclear. **PURPOSE** The purpose of this in vitro study was to measure the impact of lighting conditions on the accuracy (trueness and precision) of different IOSs. **MATERIAL AND METHODS** A typodont was digitized by using an extraoral scanner (L2i; Imetric) to obtain a reference standard tessellation language (STL) file. Three IOSs were evaluated-iTero Element, CEREC Omnicam, and TRIOS 3-with 4 lighting conditions-chair light 10 000 lux, room light 1003 lux, natural light 500 lux, and no light 0 lux. Ten digital scans per group were recorded. The STL file was used as a reference to measure the discrepancy between the digitized typodont and digital scans by using the MeshLab software program. The Kruskal-Wallis, 1-way ANOVA, and pairwise comparison were used to analyze the data. **RESULTS** Significant differences for trueness and precision mean values were observed across different IOSs tested with the same lighting conditions and across different lighting conditions for a given IOS. In all groups, precision mean values were higher than their trueness values, indicating low relative precision. **CONCLUSIONS** Ambient lighting conditions influenced the accuracy (trueness and precision) of the IOSs tested. The recommended lighting conditions depend on the IOS selected. For iTero Element, chair and room light conditions resulted in better accuracy mean values. For CEREC Omnicam, zero light resulted in better accuracy, and for TRIOS 3, room light resulted in better accuracy.

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Intraoral digital scans: Part 1: Influence of ambient scanning light conditions on the accuracy (trueness and precision) of different intraoral scanners

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Conflict of Interest

The authors did not have any conflict interest, financial or personal, in any of the materials described in this study.

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Keywords: Accuracy; Ambient light scanning conditions; Digital impression; Intraoral scanner; Prosthodontics.

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ABSTRACT

Statement of problem. Digital scans have increasingly become an alternative to conventional impressions. Although previous studies have analyzed the accuracy of the available intraoral scanners (IOSs), the effect of the light scanning conditions on the accuracy of those IOS systems remains unclear.

Purpose. The purpose of this in vitro study was to measure the impact of lighting conditions on the accuracy (trueness and precision) of different IOSs.

Material and methods. A typodont was digitized using an extraoral scanner (L2i; Imetric) to obtain a reference standard tessellation language (STL) file. Three IOSs were evaluated: iTero Element, Cerec Omnicam, and TRIOS 3 with 4 lighting conditions: chair light 10 000 lux; room light 1003 lux; natural light 500 lux; and no light 0 lux. Ten digital scans per group were recorded. The STL file was used as a reference to measure the discrepancy between the digitized typodont and digital scans using MeshLab software. The Kruskal-Wallis, 1-way analysis of variance (ANOVA), and pair-wise comparison were used to analyze the data.

Results. Significant differences for trueness and precision mean values were observed across different IOSs tested with the same lighting conditions and across different lighting conditions for a given IOS. In all groups, precision mean values were higher than their trueness values, indicating low relative precision.

Conclusions. Ambient lighting conditions influenced the accuracy (trueness and precision) of the IOSs tested. The recommended lighting conditions depend on the IOS selected. For iTero Element, chair and room light conditions resulted in better accuracy mean values. For Cerec Omnicam, zero light resulted in better accuracy, and for TRIOS 3, room light resulted in better accuracy.

CLINICAL SIGNIFICANCE

The standardization of ambient lighting conditions in private practice is essential to improving the accuracy of intraoral digital scanning based on the make and model of the scanner.

INTRODUCTION

Intraoral scanning has been commonly and successfully integrated into clinical dentistry.¹⁻⁹

Digital scanning techniques are a clinically acceptable alternative to conventional impression making for tooth and implant-supported crowns and short-span fixed dental prostheses.¹⁰⁻²¹

However, scanning accuracy has been shown to differ based on the different scanning technologies.^{10,17-30} However, these studies did not analyze how lighting conditions affect scanning accuracy. A previous study has analyzed the impact of ambient scanning light conditions on the accuracy of an intraoral scanner (IOS).²⁹ However, only a single IOS was evaluated, and the different ambient scanning light conditions in a practice environment should be considered.^{30,31}

Scanning accuracy can be affected by the scanner selected, the resolution at which the tooth is digitized, and the different fitting and smoothing algorithms that may be used to postprocess the surfaces.^{2,9-20,31} Furthermore, errors may result from the individual choices made by an operator regardless of the technology chosen, including calibration,³¹ scanning conditions,^{32,33} handling and learning,^{33,34} surface characteristics,³⁵⁻³⁸ scanning angle or scanning protocols,^{21,39,40} and the reconstruction and rendering methods used.

The accuracy of the scanner is defined in ISO 5725-1 and DIN 55350-13.^{41,42} Trueness relates to the ability of the scanner to reproduce a dental arch as close to its true form as possible

without deformation or distortion, while precision indicates the difference among images acquired by repeated scanning under the same conditions.^{12,41}

The purpose of the present in vitro study was to measure the impact of various ambient scanning light conditions on the accuracy of 3 different IOS systems. The null hypotheses were that no significant difference would be found in the digital scan accuracy (trueness and precision) of the 3 different IOSs under the 4 different ambient scanning light conditions evaluated and that no significant difference would be found in the digital scan accuracy (trueness and precision) of the 3 different IOSs under the same lighting condition.

MATERIAL AND METHODS

A dental simulator mannequin (NISSIM Type 2; Nissim) with a mandibular typodont set (Hard gingiva jaw model MIS2010-L-HD-M-32; Nissim) was used. On the selected typodont, the second right premolar was missing (Fig. 1). Three marker dots (Suremark SL-10; Suremark) were added onto the mandibular typodont to aid future superimposition and 3D measurements. The markers were attached to the occlusal surfaces of the first left molar, first right premolar, and second right molar (Fig.1B). The reference typodont was then digitized as the reference model using a structured light laboratory scanner (L2 Scanner; Imetric) to obtain a standard tessellation language (STL) file. The laboratory scanner had been previously calibrated following the manufacturer's instructions. The manufacturer of this scanner reports a trueness of <5 µm and a precision of <10 µm.

A prosthodontist (M.R.L.) with 8 years of prior experience using IOSs recorded different digital scans. In order to replicate the clinical environment, the interincisal opening was standardized to 50 mm. In addition, the mannequin was fixed on the head support of a dental

chair, and the IOSs were always positioned on the left side of the chair. Three IOSs were evaluated (Table 1) at 4 ambient light settings (Table 2).

For the chair light (CL) group, a room with a dental chair (A-dec 500; Adec) and no windows was selected. The light-emitting diode (LED) light of the chair had an intensity of 15 000 lux and 4100 K which was oriented 45 degrees at 58 cm from the mannequin. The lighting in the room was 6 fluorescent tubes of 54 W, 5000 lumens (GE F54W-T5-841-ECO; Ecolux High Output) with a white spectrum color temperature (4100 K) ceiling light and 10 000 lux measured with a light meter (LX1330B Light Meter; Dr. Meter Digital Illuminance).

For the room light (RL) group, the light of the chair was turned off, and only the ceiling light was used, with no windows or natural light. The illuminance of the room was 1003 lux as measured with the same light meter. For the natural light (NL) group, a room with natural light of 500 lux through windows as measured with the same light meter was used. For the zero light (ZL) group, a room with no light and no windows was used.

Ten digital scans per system were made for each group. The control STL file was used as a reference digital model to compare the distortion with the 120 STL files obtained. The definition of trueness in the experiment was defined as the average absolute distance between the reference model and the scanned model. The precision was defined as the distances between points of the reference model and the scanned model.^{41,42} Both trueness and precision were computed from the signed distance data according to the definitions.

For the statistical analysis of the scanned models, the software package MeshLab was used to perform the geometric preprocessing of the scanned models of the typodont, and MATLAB software was used to postprocess the data before statistical analysis. Statistical

software (IBM SPSS Statistics v25 for Windows; IBM Corp) was used to perform all statistical analysis.

The STL file format represented the scanned data as a triangle soup, such as a set of topologically nonconnected triangles, $\Delta_i = \{p_{i1}, p_{i2}, p_{i3}\}, i \in [1, n]$, that define the surface of the dental model. $p_{ij} \in \mathbb{R}^3$ was the j^{th} vertex of the i^{th} triangle ($j \in \{1,2,3\}$). This implies that each vertex on the mesh appears more than once in the triangle soup. Each scanning process resulted in a completely different set of triangles, all representing the same physical model. For this, the co-incident vertices of the triangle soup were unified to construct a topologically connected triangle mesh $M(V, F)$. Here, $V = \{v_1, \dots, v_n\}, v_i \in \mathbb{R}^3$ was the set of unified vertices and $F = \{(i, j, k)\}, i, j, k \in [1, n], i \neq j \neq k$ described the triangular faces formed by the vertices (Fig. 2A). This was performed using MeshLab.

To statistically analyze the scanned data, the primary task was to compute the spatial deviations of a treatment scanned model $S(V_S, F_S)$ with respect to the control STL model $T(V_T, F_T)$. For a vertex $v \in V_S$, the deviation was defined as the signed distance, $d_T(v)$, between v and the closest face $f \in F_T$ to v . The distance was positive if v was on the positive side of T . Mathematically, this could be computed as the sign of the dot product $\langle v - c_f, n_f \rangle$. Here, c_f and n_f were the centroid and normal of the closest face f respectively (Fig. 2B). Given a scan S , the error metric was then defined as the set $E(S) = \{d_T(v) \forall v \in V_S\}$ (Fig. 3).

For a set of multiple scanned models (S_1, \dots, S_n) from a given treatment population (such as IOS-1 group under chair lighting), the signed distance denoted as the set $E(B, L) = \cup E(S_i), i \in [1, n]$ was defined as the error distribution of the whole population. Here, B is the IOS group and L is the ambient scanning light condition.

The 2 main conditions that must hold true for computing the error in the treatment scans with respect to the control scan were as follows: both S and T were open orientable surfaces. By orientable is meant that they had 2 well-defined sides. Mathematically, this implied that all triangular faces were consistently normally oriented. Also, both S and T were geometrically aligned in 3-dimensional space.

The first condition was satisfied during the vertex unification in MeshLab. For the second condition, any given intraoral scan S was first aligned with the typodont control STL_C using the iterative-closest point algorithm. This was achieved through the following steps using the MeshLab software (Fig. 4). Firstly, a treatment scan was loaded along with the control mesh; secondly, 4 pairs of points were (approximately) chosen across the 2 meshes. Three of these 4 points were the spherical landmarks that were physically added. The fourth was a prominent crease landmark that could be easily identified. Lastly, once the correspondence was selected, the iterative closest point algorithm was applied until convergence and was repeated until the error between the aligned meshes was minimized.

One of the key issues in performing a statistical evaluation of errors was that the scanned models from different scanners resulted in distinct boundary conditions (Fig. 5). Specifically, the mesh outermost mesh vertices or, in other words, the ones that form the boundary of the surface were not aligned to the control mesh. Because of this, the signed distances of these vertices were extreme outliers that were not considered in the analysis. The challenge was that there was no deterministic rule on the basis of which these vertices could be identified. One option that was considered was to trim or crop vertices below a certain height from the dataset. However, this was rejected because of the nonlinear geometry of the typodont.

In order to mitigate this issue, statistical postprocessing was performed on each given dataset E(B, L) whereby extreme outliers were removed from the dataset before performing statistical tests (such as analysis of variance (ANOVA) and multi-comparison). The outliers were identified as error values that lie more than 3.0 times the interquartile range below the first quartile or above the third quartile.

RESULTS

For the IOS-1 group, the performance was better under the CL and RL conditions when considering the means and standard deviation of trueness and precision. For the IOS-2 group, ZL had the smallest mean and standard deviation of both trueness and precision (Table 3). For the IOS-3 group, the performance was better under NL and RL than under CL and ZL with respect to the mean and standard deviation of trueness and precision (Fig. 6).

Before conducting the ANOVA, normality testing for residuals in the ANOVA was performed using the Kolmogorov-Smirnov test. For both precision and trueness, the result showed that the data were not normally distributed. Therefore, 2-way ANOVA could not be performed on 2 datasets. Consequently, the aligned rank transform tool (ARTool)⁴³ was selected to perform the aligned rank transformation on the data, and then 2-way ANOVA was conducted on the 2 datasets. The P value of the interaction term of the IOS and ambient scanning light conditions in 2 datasets were both lower than .05, which means there was a significant interaction effect of IOS and ambient scanning light conditions on precision and trueness. Also, the P value of the main effect terms of the IOS and ambient scanning light conditions in the 2 datasets were all lower than .05, which means both factors had significant main effects on precision and trueness.

The accuracy (trueness and precision) of ambient scanning light conditions was compared for each IOS system. Because the data were not normally distributed, the Kruskal-Wallis 1-way ANOVA was conducted for ambient scanning light conditions for each IOS individually. A pair-wise comparison was also performed. The results showed that precision mean values were higher than their trueness values, which means that their relative precision was low. Moreover, by performing a pair-wise multicomparison for trueness and precision for the different IOS groups (Table 4), the effect of ambient scanning light conditions for trueness and precision were different. In the IOS-1 group, RL and NL produced significant differences in both trueness and precision. CL and NL also produced differences in both trueness and precision. However, differences in precision were only found between RL and NL, and between CL and ZL. In the IOS-2 group, significant differences in both trueness and precision were found between CL and ZL and between NL and ZL. In the IOS-3 group significant differences in both precision and trueness were found between NL and ZL and significant differences in trueness between RL and NL and between RL and CL. However, significant difference in precision were found between RL and ZL and between CL and ZL.

Comparison of accuracy (trueness and precision) was tested for each IOS system for each ambient scanning light condition evaluated. Because the data were not normally distributed, the Kruskal-Wallis 1-way analysis of variance (ANOVA) was conducted for ambient scanning light conditions for each IOS individually. A pair-wise comparison was also performed. The power of the ANOVA test indicated that the size of the datasets was adequate. For trueness, except for IOS-1 and IOS-3 under ZL, all other pairs had statistically significant differences ($P<.05$). For precision, except for IOS-1 and IO-3 under RL and CL, and IOS-1 and IOS-3 under ZL, all other pairs had statistically significant differences ($P<.05$).

DISCUSSION

Significant differences were found among the 3 IOSs systems tested under the same ambient scanning light conditions, and significant differences were found among the 4 scanning light conditions using the same IOS system; consequently, the null hypotheses were rejected. Dental studies that analyzed the impact of different ambient light conditions on the accuracy of intraoral digitizer systems are scarce.⁴² However, this scanning-based error has been analyzed previously in engineering studies.⁴⁴⁻⁴⁷

Recommendations for the optimal operating light in a dental operatory are scarce.⁴⁸⁻⁵⁰ In 1979, Viöhl⁴⁸ described as ideal 500 lux room light conditions and 2500 lux for the dental chair illumination. In 2011, the European Standard for Illumination (EN 12464) recommended 500 lux for general illumination, 1000 lux in the medical or examination rooms, and 10 000 lux for the operating cavity.⁴⁹ In the present study, the chair, room, and natural light illumination were in accordance with the recommended European Standards.

Based on the present in vitro study, ambient light conditions significantly influenced the accuracy of all IOSs tested. For iTero Element, CL and RL led to better trueness and precision mean values than the other light conditions tested; for the CEREC Omnicam, ZL scanning conditions presented the better trueness and precision mean values; and, for the TRIOS 3, RL scanning conditions produced better trueness and precision mean values. However, the NL conditions evaluated did not provide the highest accuracy when using the IOSs tested.

Scanning accuracy differences based on the different scanning technologies were identified in previous studies.^{10,18-27,41-48} Both iTero Element and TRIOS 3 IOSs scanners use the parallel confocal imaging technique.²² However, while the RL resulted in the best accuracy mean

values with both systems, iTero Element performed marginally better under CL. However, CEREC Omnicam IOS system uses a triangulation technique, with better accuracy under ZL.

The present study showed that precision mean values in all groups were higher than their trueness values, indicating that their relative precision was low. Previous studies that have analyzed the accuracy of the digital scans performed with different IOS systems,^{10-28,44-48} have not provided analysis on how lighting conditions affect scanning accuracy, which makes the accuracy values reported questionable. Additionally, the different methodology used made comparisons between the available studies difficult because of the complexity and area of the geometry analyzed (prepared tooth, sextant, or complete arch), superimposition method selected (best fit algorithm or iterative closest point algorithm), and/or reference model used.

Arakida et al²⁹ evaluated the influence of the illuminance (0, 500, and 2500 lux) and color temperature (3900, 4100, 7500, and 19 000 K) of the lighting on the accuracy of scans made with the True Definition IOS. The 500 lux and 3900 K obtained the highest accuracy, but the numerical values are not comparable with the those of the present study as different technology was used, only 2 teeth were digitized, and the reference model was an STL file obtained through a CMM machine.

The results of this study were obtained by performing a digital scan on a completely dentate arch in an in vitro environment. Evaluations of other clinical scenarios using IOSs may, however, change the outcome because of inaccuracies from edentulous areas with a higher level of nonattached tissues. Further studies are needed to fully understand the impact of lighting conditions on the accuracy of the available intraoral digitizer systems in the clinical environment.

CONCLUSIONS

With the limitations of this in vitro study, the following conclusions were drawn:

1. Lighting conditions influenced the accuracy (trueness and precision) of the digital scans performed with any of the 3 intraoral scanners tested.
2. An ideal lighting condition that resulted in best accuracy for all scanning technologies was not found.
3. Consequently, lighting condition should be selected based on the specific IOS system used.
4. For the iTero Element scanner, chair (10 000 lux) and room (1003 lux) lighting improved the trueness and precision mean values.
5. For the CEREC Omnicam scanner, zero lighting resulted in better trueness and precision mean values.
6. For the TRIOS 3 scanner, room (1003 lux) lighting provided better trueness and precision mean values.

REFERENCES

1. Duret F. Toward a new symbolism in the fabrication of prosthetic design. *Les Cahiers de Prothèse* 1985;13:65-71.
2. Zimmermann M, Mehl A, Mörmann WH, Reich S. Intraoral scanning systems - a current overview. *Int J Comput Dent* 2015;18:101-29.
3. Goracci C, Franchi L, Vichi A, Ferrari M. Accuracy, reliability, and efficiency of intraoral scanners for full-arch impressions: a systematic review of the clinical evidence. *Eur J Orthod* 2016;38:422-8.
4. Chochlidakis KM, Papaspyridakos P, Geminiani A, Chen CJ, Feng IJ, Ercoli C. Digital versus conventional impressions for fixed prosthodontics: A systematic review and meta-analysis. *J Prosthet Dent* 2016;116:184-90.
5. Mangano F, Gandolfi A, Luongo G, Logozzo S. Intraoral scanners in dentistry: A review of current literature. *BMC Oral Health* 2017;17:149-51.
6. Ahlholm P, Sipilä K, Vallittu P, Jakonen M, Kotiranta U. Digital versus conventional impressions in fixed prosthodontics: a review. *J Prosthodont* 2018;27:35-41.
7. Christensen GJ. Impressions are changing: Deciding on conventional, digital or digital plus in-office milling. *J Am Dent Assoc* 2009;140:1301-4.
8. Baheti JM, Soni UN, Gharat NV, Mahagaonkar P, Khokhani R, Dash S. Intra-oral scanners: a new eye in dentistry. *Austin J Orthopade Rheumatol* 2015;2:3-5.
9. Alghazzawi TF. Advancements in CAD/CAM technology: options for practical implementation. *J Prosthodont Res* 2016;60:72-84.
10. Patzelt SB, Vonau S, Stampf S, Att W. Assessing the feasibility and accuracy of digitizing edentulous jaws. *J Am Dent Assoc* 2013;144:914-20.

11. Flügge TV, Schlager S, Nelson K, Nahles S, Metzger MC. Precision of intraoral digital impressions with iTero and extraoral digitalization with iTero and a model scanner. *Am J Orthod Dentofacial Orthop* 2013;144:471-8.
12. Papaspyridakos P, Chen CJ, Gallucci GO, Doukoudakis A, Weber HP, Chronopoulos V. Accuracy of implant impressions for partially and completely edentulous patients: a systematic review. *Int J Oral Maxillofac Implants* 2014;29:836-45.
13. De Luca Canto G, Pachêco-Pereira C, Lagravere MO, Flores-Mir C, Major PW. Intra-arch dimensional measurement validity of laser-scanned digital dental models compared with the original plaster models: a systematic review. *Orthod Craniofac Res* 2015;18:65-76.
14. Al-Jubuori O, Azari A. An introduction to dental digitizers in dentistry. A systematic review. *J Chem Pharm Res* 2015;7:10-20.
15. Aragón ML, Pontes LF1, Bichara LM, Flores-Mir C, Normando D. Validity and reliability of intraoral scanners compared to conventional gypsum models measurements: a systematic review. *Eur J Orthod* 2016;38:429-34.
16. Tsirogiannis P, Reissmann DR, Heydecke G. Evaluation of the marginal fit of single-unit, complete-coverage ceramic restorations fabricated after digital and conventional impressions: A systematic review and meta-analysis. *J Prosthet Dent* 2016;116:328-35.
17. Joda Joda T, Zarone F, Ferrari M. The complete digital workflow in fixed prosthodontics: a systematic review. *BMC Oral Health* 2017;17:124-31.
18. Renne W, Ludlow M, Fryml J, Schurch Z, Mennito A, Kessler R, et al. Evaluation of the accuracy of 7 intraoral scanners: An in vitro analysis based on 3-dimensional comparison. *J Prosthet Dent* 2017;118:36-42.

19. Rutkūnas V, Gečiauskaitė A, Jegelevičius D, Vaitiekūnas M. Accuracy of digital implant impressions with intraoral scanners. A systematic review. *Eur J Oral Implantol* 2017;0:101-20.
20. Medina-Sotomayor P, Pascual-Moscardó A, Camps I. Relationship between resolution and accuracy of four intraoral scanners in complete-arch impressions. *J Clin Exp Dent* 2018;10:e361-6.
21. Abduo J, Elseyoufi M. Accuracy of intraoral scanners: A systematic review of influencing factors. *Eur J Prosthodont Restor Dent* 2018;26:101-21.
22. Takeuchi Y, Koizumi H, Furuchi M, Sato Y, Ohkubo C, Matsumura H. Use of digital impression systems with intraoral scanners for fabricating restorations and fixed dental prostheses. *J Oral Sci* 2018;60:1-7.
23. Tomita Y, Uechi J, Konno M, Sasamoto S, Iijima M, Mizoguchi I. Accuracy of digital models generated by conventional impression /plaster-model methods and intraoral scanning. *Dent Mater J* 2018;37:628-33.
24. Malik J, Rodriguez J, Weisbloom M, Petridis H. Comparison of accuracy between a conventional and two digital intraoral impression techniques. *Int J Prosthodont* 2018;31:107-13.
25. Nedelcu R, Olsson P, Nyström I, Rydén J, Thor A. Accuracy and precision of 3 intraoral scanners and accuracy of conventional impressions: A novel in vivo analysis method. *J Dent* 2018;69:110-8.
26. Khraishi H, Duane B. Evidence for use of intraoral scanners under clinical conditions for obtaining full-arch digital impressions is insufficient. *Evid Based Dent* 2017;18:24-5.
27. Patzelt SB, Emmanouilidi A, Stampf S, Strub JR, Att W. Accuracy of full-arch scans using intraoral scanners. *Clin Oral Investig* 2014;18:1687-94.

28. Mennito AS, Evans ZP, Lauer AW, Patel RB, Ludlow ME, Renne WG. Evaluation of the effect scan pattern has on the trueness and precision of six intraoral digital impression systems. *J Esthet Restor Dent* 2018;30:113-8.
29. Arakida T, Kanazawa M, Iwaki M, Suzuki T, Minakuchi S. Evaluating the influence of ambient light on scanning trueness, precision, and time of intra oral scanner. *J Prosthodont Res* 2018;62:324-9.
30. Logozzo S, Zanetti EM, Franceschini G, Kilpela A, Makynen A. Recent advances in dental optics - part I: 3D intraoral scanners for restorative dentistry. *Opt Lasers Eng* 2014;54:187-96.
31. Richert R, Goujat A, Venet L, Viguie G, Viennot S, Robinson P, et al. Intraoral scanners technologies: A review to make a successful impression. *J Healthc Eng* 2017:1-9.
32. Shearer BM, Cooke SB, Halenar LB, Reber SL, Plummer JE, Delson E, et al. Evaluating causes of error in landmark-based data collection using scanners. *PLOS One* 2017:1-37.
33. Kim J, Park JM, Kim M, Heo SJ, Shin IH, Kim M. Comparison of experience curves between two 3-dimensional intraoral scanners. *J Prosthet Dent* 2016;116:221-30.
34. Lim JH, Park JM, Kim M, Heo SJ, Myung JY. Comparison of digital intraoral scanner reproducibility and image trueness considering repetitive experience. *J Prosthet Dent* 2018;119:225-32.
35. Alghazzawi TF, Al-Samadani KH, Lemons J, Liu PR, Essig ME, Bartolucci AA, et al. Effect of imaging powder and CAD/CAM stone types on the marginal gap of zirconia crowns. *J Am Dent Assoc* 2015;146:111-20.
36. Anh JW, Park JM, Chun YS, Kim M, Kim M. A comparison of the precision of three-dimensional images acquired by two intraoral scanners: effects on tooth irregularities and scanning direction. *Korean J Orthod* 2016;46:3-12.

37. Müller P, Ender A, Joda T, Katsoulis J. Impact of digital intraoral scan strategies on the impression accuracy using the TRIOS pod scanner. *Quintessence Int* 2016;47:343-9.
38. Park JM. Comparative analysis on reproducibility among 5 intraoral scanners: sectional analysis according to restoration type and preparation outline form. *J Adv Prosthodont* 2016;8:354-62.
39. Carbajal Mejía JB, Wakabayashi K, Nakamura T, Yatani H. Influence of abutment tooth geometry on the accuracy of conventional and digital methods of obtaining dental impressions. *J Prosthet Dent* 2017;118:392-9.
40. Li H, Lyu P, Wang Y, Sun Y. Influence of object translucency on the scanning accuracy of a powder-free intraoral scanner: A laboratory study. *J Prosthet Dent* 2017;117:93-101.
41. International Organization for Standardization. ISO 5725-1. Accuracy (trueness and precision) of measuring methods and results. Part-I: General principles and definitions. Berlin: International Organization for Standardization; 1994. Available at:
<https://www.iso.org/standard/11833.html>
42. Ender A, Mehl A. Accuracy of complete-arch dental impressions: a new method of measuring trueness and precision. *J Prosthet Dent* 2013;109:121-8.
43. Wobbrock JO, Findlater L, Gergle D, Higgins JJ. The aligned rank transform for nonparametric factorial analyses using only anova procedures. In: *Proceedings of the 2011 Annual Conference on Human Factors in Computing Systems - CHI '11*. New York, New York, USA: ACM Press; 2011. p.143.
44. Voisin S, Fofou S, Truchetet F, Page D, Abidib M. Study of ambient light influence for three dimensional scanners based on structured light. *Opt Eng* 2007;46:030502.

45. Boehler W, Bordas VM, Marbs A. Investigating laser scanner accuracy. In: Proceedings of XIXth CIPA WG 6, International Symposium, Antalya, Turkey. 2004. p.696-702.
46. Vukašinović N, Možina J, Duhovnik J. Correlation between incident angle, measurement distance, object colour and the number of acquired points at CNC laser scanning. *J Mech Eng* 2012;58:23-8.
47. Cuesta E, Rico JC, Fernández P, Blanco D, Valino G. Influence of roughness on surface scanning by means of a laser stripe system. *Int J Adv Manuf Technol* 2009;43:1157-66.
48. Viohl J. Dental operating lights and illumination of the dental surgery. *Int Dent J* 1979;29:148-63.
49. European lightening standard EN12464-1. Light and lighting - Lighting of work places - Part 1: Indoor work places; 2011. p. 1-29.
50. International Organization for Standardization. ISO 9680. Dentistry operating lights. Geneva: International Organization for Standardization; 2014. Available at:
<https://www.iso.org/standard/39276.html>

TABLES

Table 1. Characteristics of intraoral scanning systems evaluated

GROUP	Open/Close system	Technology	Powdering	Color image	Image type
IOS-1 iTero Element (Cadent LTD)	Open	Parallel confocal microscopy technique Illuminates the surface of the object with three beams of different colored light (red, green, or blue) which combine to provide white light.	No	Yes	Photography
IOS-2 Omniscam (Cerec-Sirona)	Open	Active triangulation (Multicolor stripe projection).	No	Yes	Film (video)
IOS-3 TRIOS 3 (3Shape)	Open	Confocal microscopy technology. Ultrafast optical sectioning. Light source provides an illumination pattern to	No	Yes	Photography

		cause a light oscillation on the object.			
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Table 2. Summary of different light condition settings evaluated

LIGHT CONDITION	Chair light 10 000 lux 4100 K	Room light 1003 lux 4100 K	Windows 500 lux
CL	Yes	Yes	No
RL	No	Yes	No
NL	No	No	Yes
ZL	No	No	No

CL, chair light; NL, natural light; RL, room light; ZL, zero light.

Table 3. Statistical aggregates of error for all IOS groups (IOS-1, IOS-2, and IOS-3) against lighting conditions (CL, RL, NT, ZL). Values in micrometers.

Brand	Lighting	Precision			Trueness		
		Mean	SD	Median	Mean	SD	Median
IOS-1	CL	192.81	51.56	196.13	70.96	14.53	74.51
	NL	317.24	36.91	321.65	83.22	12.47	78.50
	RL	189.83	16.19	191.85	73.46	4.68	71.97
	ZL	333.89	40.55	352.66	84.82	12.36	88.60
IOS-2	CL	533.44	277.55	438.01	408.52	129.39	393.10
	NL	545.55	180.72	475.60	445.19	135.66	370.42
	RL	431.70	234.33	384.74	326.01	112.04	315.93
	ZL	321.02	90.59	279.79	281.84	77.12	247.06
IOS-3	CL	254.40	146.69	208.19	132.69	28.73	130.99
	NL	207.65	6.75	207.70	139.49	21.61	139.26
	RL	204.48	6.34	203.86	105.59	29.00	94.31
	ZL	324.78	245.56	216.72	118.12	57.84	92.22

CL, chair light; NL, natural light; RL, room light; SD, standard deviation; ZL, zero light.

Table 4. Power of ANOVA test of trueness and precision by IOS groups (IOS-1, IOS-2, and IOS-3) and light conditions. CL, chair light; IOS, intraoral scanner; NL, natural light; RL, room light; ZL, zero light.

Sample 1- Sample 2	CL		NL		RL		ZL	
	Trueness	Precision	Trueness	Precision	Trueness	Precision	Trueness	Precision
IOS1- IOS2	0.000	0.000	0.000	0.017	0.000	0.000	0.000	0.334
IOS1- IOS3	0.038	0.121	0.015	0.009	0.015	0.223	0.310	0.006
IOS2- IOS3	0.031	0.007	0.010	0.000	0.010	0.001	0.002	0.071

FIGURES

Figure 1. A, Dental simulator model with clinically standardized interincisal opening of 50 mm. B, Dentate typodont with mandibular right second premolar missing with 3 markers on occlusal surfaces on right first premolar and second molar typodont teeth.

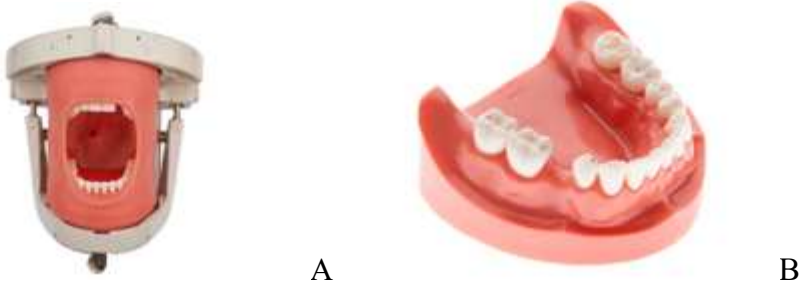


Figure 2. Geometric preliminaries for typodont scan analysis. A, Triangle soup (left) to triangle mesh (right) using vertex unification. B, Signed distance.

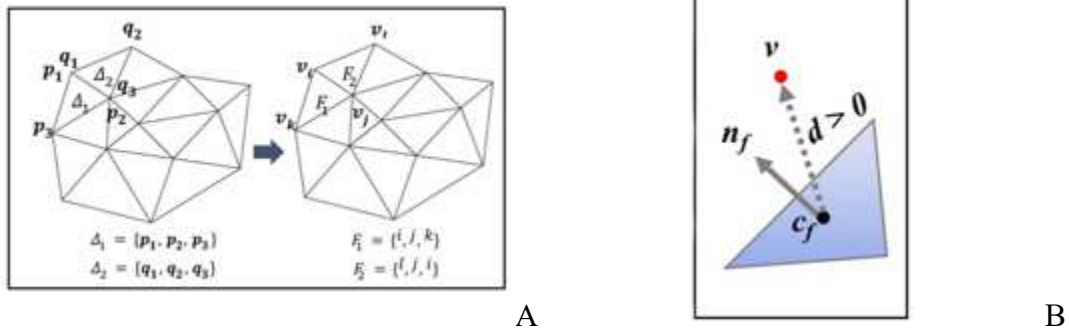


Figure 3. SEQ figure/* ARABIC 2: Color coded signed distance field for treatment scan with respect to control mesh. *Blue* color represents areas with significantly higher dimensions and *red* color areas with significantly smaller dimensions.

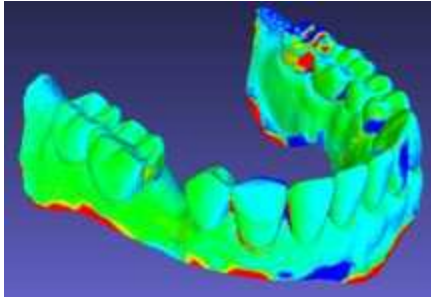


Figure 4. Typodont mesh alignment using iterative closest point algorithm in MeshLab. A, Misaligned. B, Pairs of correspondence (shown with color codes) points chosen. C, Aligned meshes after iterative closest point technique.

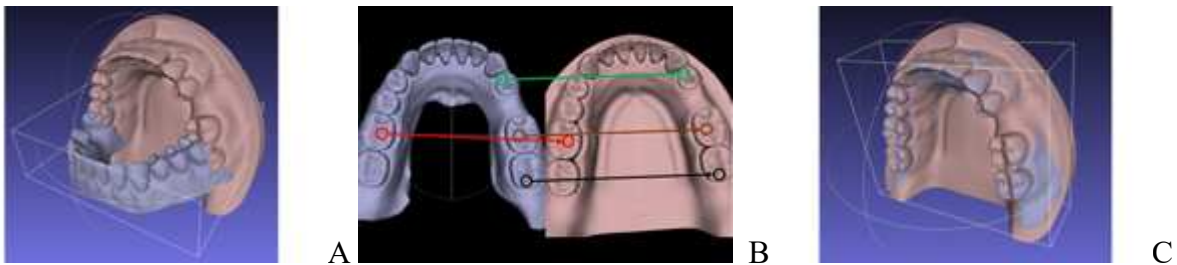


Figure 5. SEQ figure/* ARABIC 4: Extreme outliers for scanned model.

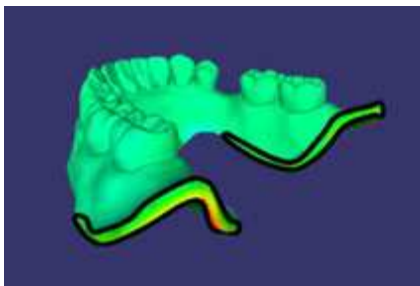


Figure 6. Boxplot of minimum, maximum, interquartile range, medians, and outliers for trueness and precision of different IOSs and ambient scanning light conditions. CL, chair light; IOS, intraoral scanner; NL, natural light; RL, room light; ZL, zero light.

